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#### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/US (22) International Filing Date: 5 June 1990 (30) Priority data: 401,613 31 August 1989 (31.08.8) (60) Parent Application or Grant (63) Related by Continuation US 401 Filed on 31 August 1989	(05.06. 9)	(75) Inventors/Applicants (for US only): ROSSI, John, J. [US/US]; 346 Cimmeron Trail, Glendora, CA 91740 (US). CHANG, Pairoj [US/US]; 949 Avenida Loma Vista, San Dimas, CA 91773 (US). KAPLAN, Bruce, E. [US/US]; 825 N. Indian Hill, Claremont, CA 91711 (US).  (74) Agent: IRONS, Edward, S.; 919 18th Street, N.W., Suite 800, Washington, DC 20006 (US).
(71) Applicant (for all designated States except US): HOPE [US/US]; 1450 East Duarte Road, E 91010-0269 (US).	CITY Juarte,	OF Published CA With international search report.

(54) Title: CHIMERIC DNA-RNA CATALYTIC SEQUENCES

#### DRDRD-1

5' GGUGCGAGAGCGUCAGUAUUAAGCGG 3' - HIV 792-817
3' CCACGCTCTCGCA TCATAATTCGCC 5'

A C UG
A
G U =RNA
C G C
A T
G C
G C
A G
G T

(57) Abstract

This invention provides chimeric DNA/RNA catalytic molecules useful to cleave RNA sequences. The invention specifically provides two different chimeric DNA-RNA-DNA-RNA-DNA catalytic molecules which are targeted to cleave HIV-I RNA sequences. These chimeric molecules include DNA sequences which flank a catalytic RNA center. Interaction with the HIV-I substrate RNAs is achieved by Watson-Crick base pairing of the DNA flanking sequences with HIV-I RNA. The catalytic ribonucleotide center cleaves the phosphodiester bond of the substrate HIV-! RNA at the expected location.

<sup>\*</sup> See back of page

#### Summary of the Invention

This invention provides chimeric DNA/RNA catalytic molecules useful to cleave RNA sequences. The invention specifically provides two different chimeric DNA-RNA-DNA-RNA-DNA catalytic molecules which are targeted to cleave HIV-1 RNA sequences. These chimeric molecules include DNA sequences which flank a catalytic RNA center. Interaction with the HIV-1 substrate RNAs is achieved by Watson-Crick base pairing of the DNA flanking sequences with HIV-1 RNA. The catalytic ribonucleotide center cleaves the phosphodiester bond of the substrate HIV-1 RNA at the expected location.

### General Description of the Invention

In general the catalytic molecules of the invention function as hammerhead or hairpin ribozymes. The preferred molecular construct consists of two known RNA catalytic sequences each flanked by a DNA sequence at the respective 3' and 5' termini and coupled by a DNA sequence at the corrseponding 5' and 3' termini. These molecules may accordingly be represented by the formulae I and II:: I.

3' X - AAAG - Y - AGUAGUC - Z 5'

.3' X - CAAAG - Y - AGUAGUC - Z 5' in which X, Y and Z are DNA sequences and AAAG, CAAAG and AGUAGUC are catalytic RNA sequences.

The flanking X and Z components may be any DNA sequences that allow base pairing with the substrate RNA at appropriate positions adjacent to the substrate cleavage site. These flanking sequences may be phosphodiester, phosphorothicate, methyl phosphonate, methyl phosphate or similar moieties.

Y may be any DNA sequence that base pairs inter se in the manner required for catalytic cleavage of

1300

the substrate by the RNA sequences preferably as shown in base paired form in Formula III:

III. 5' C-G 3'
A-T
G-C
G-C
A G

The catalytic molecules of this invention can be synthesized in known manner by commercially available DNA synthesizers such as those produced by Applied Biosystems or Milligen. See, e.g., Perreault, et al, supra.

The X and Z sequences may be substituted at the respective 3' and 5' ends with ligands to facilitate cell entry, targeting within the cell and ultimate stability of the catalysts. Such ligands include by way of example but not of limitation: other nuclotides, proteins, carbohydrates, lipids, steroid hormones and cholesterol.

The catalytic molecules of the invention are administered by known and available delivery agents or systems, including, but not limited to, liposomes, defective viral particles, viral capids, and standard DNA/RNA transfective procedures.

#### Description of the Figures

Figure 1 illustrates one catalytic molecule of the invention base paired to an HIV-1 sequence. The RNA portion of the molecule is encircled.

Figure 2 illustrates a second catalytic molecule of the invention base paired to another HIV-1 sequence. The RNA portion of the molecule is encircled.

Figure 3A depicts a ribonuclease A digestion of the catalytic molecule of Figure 1 as compared with an equivalent all DNA molecule. The conditions were 10 units of commercial (Sigma) pancreatic ribonuclease in 2XSSC buffer added to the oligonucleotides which were in 10 microliters of 50 mM Tric-HC1 buffer (pH 8.0). The RNAse was incubated with the sample for 10 minutes before the 32-p end labelled DRDRD or DNA molecules were electrophoresed in a 15t polyacrylamide gel containing 8M urea. The gel was autoradiographed for 10 minutes to get the exposure depicted.

Figure 3B depicts a cleavage reaction involving the catalytic molecule of Figure 1 under conditions described in Chang, et al., Clinical Biotechnology, 2:23-31 (1990).

#### EXAMPLE I

The catalytic molecule of Figure 1 was synthesized in known manner utilizing an automated oligonucleotide synthesizer manufactured by Applied Biosystems, Inc.

The result of ribonuclease A digestion of the catalytic molecule is shown by Figure 3A.

The catalytic molecule produced, as described, was used to cleave each of a 610 nuleotide long (S-610) and a 170 nucleotide long HIV-1 gag transcript. In brief, the buffer was 50 mM Tris-HCl, pH 7.5, lmM EDTA, 10mM MgCl<sub>2</sub> at approximately 1 pmole of target, 3 pmole of ribozyme or DNA. The reactions were carried out at 37°C. for 12 hours. The substrate was either a 610 nucleotide long HIV-1 gag containing transcript (S-610) or a 172 nucleotide long HIV-1 gag containing transcript (S-610). The 5° cleavage product is indicated for both.

In Figure 3B the 5' cleavage product is shown for both transcripts. The 3' cleavage product for the 610 target is not visible due to poor reproduction of

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Specific cleavage of an HIV-1 5' LTR splice site with a similar catalytic molecule has also been obtained.

<u> 4</u>.

#### CLAIMS

 A catalytic molecule capable of cleaving an HIV-1 RNA sequence at a known ribozyme cleavage site said molecule having the formula

3' X - AAAG - Y - AGUAAGUC - Z 5'

or

3' X - CAAAG - Y - AGUAAGUC - Z 5' in which X and Z are DNA sequences that base pair with an RNA substrate at positions juxtaposed to said known cleavage site,

AAAG, CAAAG and AGUAGUC are RNA sequences,

Y is a DNA sequence that base pairs <u>inter se</u> in a manner required to permit said RNA sequences to cleave said substrate at said cleavage site.

- 2. The catalytic molecule shown by Figure 1.
- 3. The catalytic molecule shown by Figure 2.
- 4. A catalytic molecule, as defined by Claim 1, in which said RNA sequence is an HIV-1 sequence.
- 5. A catalytic molecule, as defined by Claim 4, in which said HIV-1 sequence is the HIV-1 sequence shown by Figure 1.
- 6. A catalytic molecule, as defined by Claim 4, in which the HIV-1 sequence is the HIV-1 sequence shown by Figure 2.
- 7. A catalytic molecule capable of cleaving an RNA sequence, said molecule having catalytic RNA moieties linked to first and second DNA moieties which base pair with the substrate RNA sequences flanking the cleavage site and interconnected by a third DNA sequence which base pairs <u>inter</u> se to facilitate said cleavage.

### FIG. 1 DRDRD-1

5' GGUGCGAGAGCGUCAGUAUUAAGCGG 3' - HIV 792-817
CCACGCTCTCGCA) TCATAATTCGCC 5'

A C UG
A G
C G G C
G C
G C
G G T

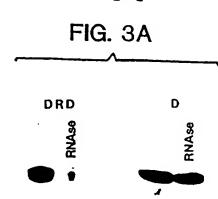
# FIG. 2 DRDRD #2

5'CGACUGGUGAGUACGCCAAAA 3' - HIV LTR 737-757
3'GCTGACCTCTCA GCGGTTTT 5'

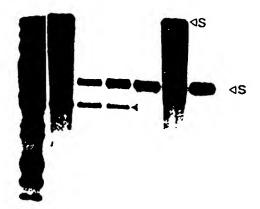
A C U G
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### INTERNATIONAL SEARCH REPORT

International Application No PCT/US90/03102

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III. DOCU	MENTS CONSIDERED TO BE RELEVANT "		
Category *	Citation of Document, 14 with Indication, where approp	riate, of the relevant passages 11	Relevant to Claim No. 14
A,P	Chemical Abstract, Volume 112, No. 12 February 1990 (Columbus, Chio, W. Gerlach, et al, "Synthetic Rif Inactivation of Prokaryotic or Extranscripts", See pages 336-337, abstract No. 51284j, Eur. Pat. Ap 21 June 1989.	, U.S.A.) cozymes for <u>in ViVo</u> ukaryotic RNA column 2, See the	1 - 7
A,P	Chemical Abstract, Volume 112, No. 07 May 1990 (Columbus, Chio, U.S. "Ribozymes as Potential Anti-HIV. Agents", See page 420, column 2, No. 17548q, Science, 1990, 247 (	.A.) N. Sarver, et al., -1 Therapeutic See the abstract	1 - 7
A,P	Chemical Abstract, Volume 112, N 12 February 1990 (Columbus, Chio M. Cotten, et al, "Ribozyme Medi RNA in Vivo", See page 501, colu abstract No. 52942j, EMBO J, 199	, U.S.A.), ated Destruction of mn 1, See the	1 - 7
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III DOCUM	The Annual Application in a	PCT/US90/03102
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reledad.	Citation of Document, 44 with Indication, where appropriate, of the relevant passages 17	Relevant to Claim No
A,P	Nature, volume 344, issued 05 April 1990, J. Peneault, et al., Mixed Deoxyribo - and Ribooligonucleotides with Catalytic activity see pages 565-567.	1-7
A,P	Proceeding of the National Academy of Sciences, Volume 86, no. 23, issued December 1989 (U.S.A.) F.H. Cameron, et al., 'Specific Gene Suppression by Engineered Ribozymes in Monkey Cells', see pages 9139 - 9143.	
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A	Chemical Abstracts, Volume 110, No. 21, issued 22 May 1989, (Columbus, Ohio, U.S.A.) T. R. Cech et al., "RNA Ribozyme Polymerases, Dephosphorylases, Restriction Endoribonucleases and Methods for Their Manufacture", See page 226, column 2, See the abstract No. 187321K, PCT Int. Appl. W08804,300 16 June 1988.	1 - 7
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